

Cleaning MyoDish chambers and electrodes

Important note: When handling the chambers, be careful and do not overbend the spring wire and to not touch the magnet at the end of the spring wire !

Do not aim the water jet on the spring wire or magnet!

DO NOT AUTOCLAVE the chambers!

1. Prepare glass container with 0.5 - 1 L of DI water
2. Remove medium from the chambers with a pipette and discard medium as biological waste
3. Remove electrodes and put them into the DI water
4. Unscrew the sensor boards
 - a. check them for any defects and medium / fluid remnants
 - b. clean them with a clean cotton swab and a little DI water and let dry thoroughly
 - c. when dry and clean: place the sensor boards carefully into antistatic bag and keep at a dry, clean place at room temperature
5. Put the chambers carefully into the DI water (together with the electrodes)
6. Remove the DI water, add 1 L of fresh DI water + a few drops of detergent with enzymes breaking down proteins, lipids and sugars (dish liquid)
7. Place on horizontal rocker for 1-2 hours or overnight (gentle rocking!)
8. Rinse thoroughly with water until detergent is fully washed away
9. Add 400 ml of ethanol / isopropanol + 400 ml of DI water and put on rocker for another 1-2 hours or overnight (gentle rocking!)
10. Rinse with DI water and let chambers dry on paper towels with the opening facing downwards in a clean environment
 - a. When dry, put them into a box and keep at a dry, clean place
11. Additional cleaning of electrodes (to remove drug remnants etc.): put them into a 100ml glass bottle, add 50-60 ml of 100% isopropanol and incubate for 1-2 hours or overnight with gentle rocking
12. Remove isopropanol, let remnants of isopropanol evaporate and keep electrodes in a closed glass container
13. If possible: autoclave electrodes before next use